



The name of university: Program name The course name: Course code: Academic year / level:

University of Benghazi Molecular diagnostics Clinical Biochemistry MLSC-201 2023-2024. Second year

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QAPA.COUR.SP.E.2a

1- Basic information:

The course name:	Clinical Biochemistry
Course coordinator	
Program (s) on which the course is given:	Molecular diagnostics
Teaching hours	4 hours/week
Language	English
Academic year / level:	2023-2024. Second year
Course approval date	

1.1 hours per week

Lecture: (2 hours/ week)

Practical: (2 hours/ week) Training: (-) Total: (hours 96)

2- Aims of Course:

- a. The course aims to provide students with a comprehensive understanding of the biochemical principles underlying human disease and to develop their knowledge and skills in clinical laboratory science, including the ability to interpret laboratory data.
- b. This course covers a wide range of topics including basic analytical techniques, metabolism and disease, endocrine and reproductive systems, renal and liver function, and therapeutic drug monitoring.
- c. It also provides fundamental knowledge of the structure and function of biomolecules and their role in human physiology, health, and disease.
- d. Through a combination of lectures, practical classes, and case-based learning, students will acquire a solid foundation in the principles and practices of clinical chemistry, which are essential for a career in clinical laboratory science, medicine, or related fields.

3- Intended Learning Outcomes of Course (ILOs):

a- Knowledge and Understanding:

After completing this course, students should be able to demonstrate:

- 1. Understand the biochemical basis of human disease.
- 2. Stare the principles of clinical laboratory science.
- 3. Underline the important biochemical analyses used in clinical biochemistry.

b- Intellectual Skills:

By the end of the course, student should be able to:

- 1. Critically evaluate and interpret biochemical data.
- 2. Apply knowledge from different areas of biochemistry and biomedical sciences.

c- Professional and Practical Skills:

By the end of the course, student should be able to:

1. Pro laboratory tests in clinical biochemistry.

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- 2. The ability to use standard operating procedures and quality control measures in the clinical laboratory.
- 3. Demonstrate knowledge of the metabolic pathways and biochemical processes involved in the diagnosis and management of diseases.
- 4. Understand the roles of various organs and organ systems in the regulation of biochemical homeostasis.
- 5. The ability to interpret laboratory data in the context of patient care.

d- General Skills:

By the end of the course, student should be able to:

- 1. Improve teamwork skills
- 2. Enhance communicate scientific ideas effectively in written and oral formats.
- 3. Develop problem-solving skills.
- 4. Develop effective time management and organizational skills.

4- Course Contents:

Tania	Hauna	Lasturas	Lab.	Practical/
Торіс	nours	Lectures		small groups
Introduction to Clinical Biochemistry				
Biomolecules and their Properties				
Metabolism				
Enzymes and Their Kinetics				
Carbohydrate Metabolism and Diabetes				
Lipid Metabolism and Cardiovascular				
Disease				
Protein Metabolism and Renal Function				
Endocrine and Reproductive Systems				
Hormones and Signaling				
Biochemical Techniques				
Diagnostic Techniques in Clinical				
Biochemistry				
Acid-Base Balance and Electrolytes				
Hematology and Coagulation Disorders				
Clinical Enzymology and Analytical				
Techniques				
Trace Elements and Minerals				
Iron Metabolism				
Inborn Errors of Metabolism				
Total.				

:Learning Methods .5

a. Lectures

- b. Tutorial
- c. Lab.

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6. Assessment methods:

Assessment Type	Date	%				
First assessment	16 th week of first term.	<mark>30</mark>				
Lab. Exam	Before the final examination	20				
Final examination (Written,)	At the end of term	<mark>50</mark>				
Total						

7. List of References:

Title	Publisher	copy	authors	Avail
				able
				place
Clinical Guide to	Philadelphia, PA: W.B.	4th	Tietz NW, Pruden EL,	
Laboratory Tests.	Saunders Company.	ed.	Siggaard-Andersen O.	
Clinical Chemistry:	London, UK: CRC Press.	2nd	Marshall WJ, Bangert	
Principles and Practice.		ed.	SK.	

8. Facilities Required for Teaching and Learning:

	Facilities Required	Notes
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Course Coordinators

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Date:...../...../...../

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Head of Department

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The name of university: Programme name: The course name: Course code: Academic year / level: University of Benghazi Molecular diagnostics Clinical Biochemistry MLSC-201 2023-2024. Second year

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1. Basic information:

	-
the course name:	Clinical Biochemistry
Course coordinator	
Programme (s) on which the course is given:	Molecular diagnostics
Teaching hours	5 hours/week
Language	English
Academic year / level:	2023-2024. Second year
Course approval date	

1.1 hours per week

Lecture: (3 hours/week)

Practical: (2 hours/week) Training: (-) Total: (hours 160)

2. Aims of Course:

- a. To provide to provide students with a comprehensive understanding of the biochemical principles underlying human disease.
- b. To develop their knowledge and skills in clinical laboratory science, including the ability to interpret laboratory data.
- c. To covers a wide range of topics including basic analytical techniques, metabolism and disease, endocrine and reproductive systems, renal and liver function, and therapeutic drug monitoring.
- d. To provides fundamental knowledge of the structure and function of biomolecules and their role in human physiology, health, and disease. Through a combination of lectures, practical classes, and case-based learning, students will acquire a solid foundation in the principles and practices of clinical chemistry, which are essential for a career in clinical laboratory science, medicine, or related fields.

3. Intended Learning Outcomes of Course (ILOs):

a- Knowledge and Understanding:

By the end of the course, student should be able to :

- 1. Understand biochemical basis of human disease
- 2 Outline of the principles of clinical laboratory science
- 3. Describe most important biochemical analyses used in clinical biochemistry

b- Intellectual Skills:

By the end of the course, student should be able to :

- 1. Evaluate and interpret biochemical data.
- 2. Illustrate knowledge from different areas of biochemistry and biomedical sciences

c- Professional and Practical Skills:

By the end of the course, student should be able to :

c1. Operate laboratory tests in clinical biochemistry

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c2. Use standard operating procedures and quality control measures in the clinical laboratory.

c3. Distinguish the metabolic pathways and biochemical processes involved in the diagnosis and management of diseases

c4. Indicate the roles of various organs and organ systems in the regulation of biochemical homeostasis

5. Interpret laboratory data in the context of patient care

d- General Skills:

- By the end of the course, student should be able to:
- d1. Develop team-work skills.
- d2. Develop communication skills.
- d3. Develop analytical and problem-solving skills in a range of settings.
- d4. Develop time-management.

4- Course Contents:

Торіс	Hours	Lectures	Lab.	Practical/ small groups
Introduction to Clinical Biochemistry				
Biomolecules and their Properties				
Metabolism				
Enzymes and Their Kinetics				
Carbohydrate Metabolism and Diabetes				
Lipid Metabolism and Cardiovascular Disease				
Protein Metabolism and Renal Function				
Liver Function and Hepatobiliary Diseases				
Endocrine and Reproductive Systems				
Hormones and Signaling				
Biochemical Techniques				
Diagnostic Techniques in Clinical Biochemistry				
Acid-Base Balance and Electrolytes				
Hematology and Coagulation Disorders				
Clinical Enzymology and Analytical Techniques				
Trace Elements and Minerals				
Iron Metabolism				
Inborn Errors of Metabolism				
Introduction to Clinical Biochemistry				
Biomolecules and their Properties				
Metabolism				
Total				

5. Learning Methods:

5 Page 3 of

- a. Lectures
- b. Tutorial
- c. Lab.

6. Assessment methods:

	Assessment Type	Date	%					
1.	First assessment	16 th week of first term.	30					
2.	Lab. Exam	Before the final examination	20					
3.	Final examination (Written,)	At the end of term	50					
	Total							

7. List of References:

Title	Publisher	сору	authors	Available place
Clinical Guide to	Saunders	4th ed.	Tietz NW,	
Laboratory Tests.	Company		Pruden EL,	
Philadelphia, PA:			Siggaard-	
W.B.			Andersen O.	
Clinical Chemistry:	London, UK:	. 2nd ed.	Marshall WJ,	
Principles and	CRC Press;		Bangert SK.	
Practice			_	

8. Facilities Required for Teaching and Learning:

	Facilities Required	Notes
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Course Coordinators

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Head of Department

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The name of university: Program name The course name: Course code: Academic year / level: University of Benghazi Molecular diagnostics Integrative Physiology BMSC-201 2023-2024. Second year

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1- Basic information:

the course name:	Integrative Physiology
Course coordinator	
Program (s) on which the course is given:	Molecular diagnostics
Teaching hours	4 hours/week
Language	English
Academic year / level:	2023-2024. Second year
Course approval date	

1.1 hours per week

Lecture: (2 hours/ week)

Practical: (2 hours/ week) Training: (-) Total: (hours 96)

2- Aims of Course:

- a. The overall aim of this course is to provide students with a comprehensive understanding of the fundamental principles of physiology and the role of different physiological systems in maintaining health and responding to environmental challenges.
- b. By the end of the course, students should have the ability to critically analyze the function of different physiological systems and their interactions, and apply this knowledge to the study of human health and disease.

3- Intended Learning Outcomes of Course (ILOs):

a- Knowledge and Understanding:

After completing this course, students should be able to demonstrate:

- 1. Demonstrate an understanding of the fundamental principles of physiology and homeostasis.
- 2. Underline the organization and function of different physiological systems in the body.
- 3. Outline the mechanisms that control cellular communication and signaling pathways.
- 4. Understand the role of different physiological systems in maintaining homeostasis and responding to environmental challenges.
- 5. Describe the function of the reproductive system and its hormonal regulation.
- 6. Demonstrate the ability to design and conduct laboratory experiments in physiology.

b- Intellectual Skills:

By the end of the course, student should be able to:

- 1. Analysis and evaluation of physiological concepts and mechanisms.
- 2. Synthesis of complex physiological systems and their interactions.
- 3. Application of physiological principles to human health and disease.

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4. Evaluate of experimental design and interpretation of results

c- Professional and Practical Skills:

By the end of the course, student should be able to:

1. Interpret experimental data and draw appropriate conclusions.

d- General Skills:

By the end of the course, student should be able to:

- 1. Improve teamwork skills
- 2. Enhance communicate scientific ideas effectively in written and oral formats.
- 3. Develop effective time management and organizational skills.
- 4. Improve Critical thinking and problem-solving

4- Course Contents:

Торіс	Hours	Lecture s	Lab.	Practical/ small groups
Introduction to Physiology and Homeostasis				
Cellular Communication and Signaling				
Pathways				
Membrane Transport and Energy Metabolis				
Nervous System and Sensory Physiology				
Cardiovascular System and Blood Pressure				
Regulation				
Respiratory System and Gas Exchange				
Renal System and Water Balance				
Digestive System and Nutrient Absorption				
Endocrine System and Hormonal Regulation				
Environmental Physiology and Adaptations to				
Stressors				
Genetic Basis of Physiology				
Exercise Physiology and Metabolic				
Adaptations				
Aging and Physiological Changes				
Reproductive System and Hormonal				
Regulation				
Immunology and Inflammation				
Pharmacology and Drug Effects on Physiology				
Body Composition and Energy Balance				
Circadian Rhythms and Biological Clocks				
Neural Control of				
Cardiovascular and				

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Tonic	Hours	Lecture	Lab.	Practical/
Торк	liours	S		small groups
Respiratory Systems				
Environmental Factors and Physiology				
Hormonal Control of Digestive System				
Mechanisms of Pain and Pain Management				
Brain and Behavior				
Autonomic Nervous System and Stress				
Response				
Reproductive Health and Fertility				
Physiology of Sleep and Sleep Disorders				
Total.				

5. Learning Methods:

- a. Lectures
- b. Tutorial
- c. Lab.

6. Assessment methods:

Assessment Type	Date	%
First assessment	16 th week of first term.	<mark>30</mark>
Lab. Exam	Before the final examination	20
Final examination (Written,)	At the end of term	50
T T	otal	100

7. List of References:

Title	Publisher	Сору	authors	Available
				place
Human Physiology:	Pearson Education	7th ed.	Silverthorn, Dee	
An Integrated	Limited		Unglaub.	
Approach.				
Textbook of Medical	Elsevier	14th ed.	Hall, John E. Guyton	
Physiology.			and Hall	

8. Facilities Required for Teaching and Learning:

	Facilities Required	Notes
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Course Coordinators

Head of Department

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The name of university: Programme name: The course name: Course code: Academic year / level: University of Benghazi Molecular diagnostics Biological Basis of Disease MOLD-201 2023-2024. Second year

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1. Basic information:

the course name:	Biological Basis of Disease
Course coordinator	
Programme (s) on which the course is given:	Molecular diagnostics
Teaching hours	5 hours/week
Language	English
Academic year / level:	2023-2024. Second year
Course approval date	

1.1 hours per week

Lecture: (3 hours/week)

Practical: (2 hours/week) Training: (-) Total: (hours 160)

2. Aims of Course:

- a. To provide students with a comprehensive understanding of the biological mechanisms underlying disease development, progression, and treatment.
- b. developing a strong foundation in the fundamental principles of pathology, immunology, microbiology, pharmacology, and genetics.

3. Intended Learning Outcomes of Course (ILOs):

a- Knowledge and Understanding:

By the end of the course, student should be able to :

a1. An in-depth understanding of the biological basis of disease.

a2. The fundamental principles of pathology, including the concept of disease, etiology, pathogenesis, and morphological changes associated with diseases.

a3. The cellular responses to injury and the molecular mechanisms that underlie these responses.

a4. The role of inflammation and immunity in the pathogenesis of diseases.

a5. The principles of neoplasia, including the molecular mechanisms that underlie the development and progression of cancer.

a6. The principles of systemic pathology, including the major diseases affecting each organ system.

b- Intellectual Skills:

By the end of the course, student should be able to :

b1. Analyze and interpret pathological data from clinical and laboratory investigations.

b2. Apply knowledge of pathological principles to diagnose and manage diseases.

b3. Critically evaluate scientific literature and research related to pathology.

b4. Design and execute experiments related to pathology.

c- Professional and Practical Skills:

By the end of the course, student should be able to :

7 Page 2 of

c1. Work effectively communicate pathological concepts to healthcare professionals and patients.

c2. Work collaboratively with healthcare professionals in the diagnosis and management of diseases.c3. Use laboratory investigations to diagnose and manage diseases.

c4. Critically evaluate laboratory investigations and their role in the diagnosis and management of diseases.

d- General Skills:

By the end of the course, student should be able to:

d1. Develop effective communication skills with diverse audience.

d2. Enhance the ability to work independently and as part of a team to achieve common goals.

d3. Improve problem-solving skills to address complex issues.

4- Course Contents:

Topic	Hours	Lectures	Lab.	Practical/
				small groups
Fundamentals of Pathology:				
Cellular and Molecular Basis of Disease:				
:Immunology and Disease				
Microbiology and Disease:				
Genetics and Genomics in Disease:				
Cardiovascular Diseases:				
Respiratory Diseases:				
Gastrointestinal Diseases:				
Renal and Urinary Tract Diseases:				
Endocrine and Metabolic Diseases				
Neurological Diseases:				
Musculoskeletal Diseases:				
Oncology:				
Infectious Diseases:				
Hematological Diseases:				
:Laboratory Safety				
Biomarkers and Diagnostics:				
Biological Basis of Disease:				
Immune System Disorders:				
Genetic Disorders:				
Neurological Disorders:				
Dermatological Disorders:				
Introduction to Pathology:				
Cellular Responses to Injury:				
Acute Inflammation:				
Chronic Inflammation:				
Tissue Repair				

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Tonic	Hours	Lectures	Lab.	Practical/
Topic	mours	Lectures		small groups
and Regeneration:				
Hemodynamic Disorders:				
Genetic and Developmental Disorders:				
Environmental Pathology:				
Immunopathology:				
Neoplasia:				
Infectious Diseases:				
Nutritional and Metabolic Diseases:				
Endocrine Pathology:				
Cardiovascular Pathology:				
Respiratory Pathology:				
Gastrointestinal Pathology:				
Total				

5. Learning Methods:

- a. Lectures
- b. Tutorial
- c. Lab.

6. Assessment methods:

	Assessment Type	Date	%			
1.	First assessment	16 th week of first term.	30			
2.	Lab. Exam	Before the final examination	20			
3.	Final examination (Written,)	At the end of term	50			
Total						

7. List of References:

Title	Publisher	сору	authors	Available place
Robbins basic	Elsevier Health		Kumar, V.,	
pathology.	Sciences.		Abbas, A. K.,	
			Aster, J. C., &	
			Robbins, S. L.	
Robbins Pathologic	Philadelphia,		Cotran, R.S.,	
Basis of Disease.	PA: Saunders.		Kumar, V.,	
			Collins, T.	

8. Facilities Required for Teaching and Learning:

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	Facilities Required	Notes
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Course Coordinators

Head of Department

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The name of university: Programme name: The course name: Course code: Academic year / level: University of Benghazi Molecular diagnostics Lab Management and Medical Informatics LMBT-202 2023-2024. Second year

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1. Basic information:

the course name:	Lab Management and Medical Informatics
Course coordinator	
Programme (s) on which the course is given:	Molecular diagnostics
Teaching hours	2 hours/week
Language	English
Academic year / level:	2023-2024. Second year
Course approval date	

1.1 hours per week

Lecture: (2 hours/week)

Practical: (hours/week) Training: (-) Total: (hours 64)

2. Aims of Course:

- a. To equip students with the necessary skills and knowledge to effectively manage and maintain medical laboratories, as well as to apply medical informatics to enhance patient care.
- b. To gain a comprehensive understanding of laboratory management, regulatory compliance, data management, and medical informatics. Ultimately, the course aims to prepare students to manage modern medical laboratories and apply medical informatics to improve patient care.

3. Intended Learning Outcomes of Course (ILOs):

a- Knowledge and Understanding:

By the end of the course, student should be able to :

1. Understand the fundamental principles of laboratory operations and management.

2. Understand the regulatory requirements governing laboratory operations and compliance.

3. List the principles of quality assurance and quality control in laboratory operations.

4. Describe the importance of data management and medical informatics in modern healthcare.

b- Intellectual Skills:

By the end of the course, student should be able to :

1. Analyze and evaluate laboratory operations to identify areas for improvement.

2. Evaluate and apply laboratory regulations and compliance requirements to ensure laboratory operations meet the necessary standards.

3. Develop and implement quality control measures to ensure accurate and reliable laboratory results.

4. Analyze and interpret medical data to provide valuable insights for patient care.

c- Professional and Practical Skills:

7 Page 2 of

By the end of the course, student should be able to :

1. Practice laboratory policies and procedures to ensure effective and efficient laboratory operations.

2. Prepare data and medical informatics strategies to improve patient care.

3. Manipulate laboratory findings and medical information to healthcare professionals and patients.

4. Modify strategies to ensure laboratory safety and compliance.

d- General Skills:

By the end of the course, student should be able to:

1. Develop problem-solving skills and critical thinking skills.

2.Develop communication skills for effective communication with healthcare professionals and patients.

3. Develop skills for managing and leading laboratory staff.

4- Course Contents:

Tonio	Houng	Lasturas	Lab.	Practical/
Горіс	nours	Lectures		small groups
Introduction to Laboratory Operations and				
Management				
Laboratory Regulations and Compliance				
Introduction to Laboratory Operations and				
Management				
Laboratory Regulations and Compliance				
Medical Informatics				
Laboratory Equipment Management and				
Maintenance				
Laboratory Safety and Compliance				
Laboratory Accreditation and Certification				
Laboratory Budgeting and Finance				
Laboratory Auditing and Inspection				
Laboratory Project Management				
Laboratory Ethics and Professionalism				
Data Analysis and Reporting				
Medical Informatics Applications in				
Healthcare				
Leadership and Team Management in the				
Laboratory				
Laboratory Waste Management				
Laboratory Personnel Management				
Laboratory Inventory Control				
Laboratory Informatics and Analytics				
Laboratory Automation and Robotics				
Total				

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:Learning Methods .5

- a. Lectures
- b. Tutorial

6. Assessment methods:

	Assessment Type	Date	%				
1.	First assessment	16 th week of first term.	40				
2.	Final examination (Written,)	At the end of term	60				
Total 1							

7. List of References:

Title	Publisher	сору	authors	Available place
Laboratory	Valenstein PN.	3rd ed.		
management				
principles and				
processes.				
Washington:				
American Society				
for Clinical				
Pathology Press				
Laboratory waste	Royal Society		O'Connor J.	
management: a	of Chemistry			
guidebook. London:				
Use of electronic	N Engl J Med.		Jha AK,	
health records in			DesRoches	
U.S. hospitals.			CM, Campbell	
			EG, Donelan	
			K, Rao SR,	
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8. Facilities Required for Teaching and Learning:

	Facilities Required	Notes
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Course Coordinators

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Head of Department

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7 Page 7 of





The name of university: Program name The course name: Course code: Academic year / level: University of Benghazi Molecular diagnostics Cell and Molecular Biology MOLD-203 2023-2024. Second year

5 Page 1 of

1. Basic information:

The course name:	Cell and Molecular Biology
Course coordinator	
Program (s) on which the course is given:	Molecular diagnostics
Teaching hours	3 hours/week
Language	English
Academic year / level:	2023-2024. Second year
Course approval date	

1.1 hours per week

Lecture: (1 hours/week)

Practical: (2 hours/week) Training: (-) Total: (hours 96)

2. Aims of Course:

- a. The course aims to provide students with a fundamental understanding of the molecular and cellular processes that underpin human health and disease.
- b. The course covers a range of topics, from the basic structure and function of cells to advanced concepts such as gene expression and signaling pathways. It also includes practical sessions designed to provide students with hands-on experience in laboratory techniques commonly used in molecular and cellular biology.

3. Intended Learning Outcomes of Course (ILOs):

a- Knowledge and Understanding:

After completing this course, students should be able to demonstrate:

- 1. Understand the basic structure and function of cells and the molecular components that underpin cellular processes.
- 2. Outline the molecular genetics, including DNA replication, transcription, translation, and gene regulation.
- 3. Describe the importance of protein structure and function in cellular processes, including enzymatic catalysis and molecular signaling.
- 4. Identify the major signaling pathways involved in cellular communication and their role in normal and disease states.

b- Intellectual Skills:

By the end of the course, student should be able to :

- 1. Critically evaluate scientific literature and experimental data.
- 2. Design and execute experimental protocols in molecular and cellular biology.
- 3. Analyze and interpret data obtained from experiments.
- 4. Apply the ability to hypotheses and design experiments to test them.

c- Professional and Practical Skills:

By the end of the course, student should be able to:

5 Page 2 of

- 1. Use laboratory equipment and perform basic techniques in molecular and cellular biology.
- 2. Work effectively as part of a team in a laboratory setting.
- 3. Interpret data obtained from experiments and present findings in written and oral formats.
- 4. Conduct research in a responsible and ethical manner.

d- General Skills:

By the end of the course, student should be able to:

- 1. Develop effective communication skills.
- 2. Improve time-management and organizational skills.
- 3. Develop problem-solving and analytical skills.
- 4. Enhance teamwork skills in a laboratory setting

4- Course Contents:

Tonia	Hours	Looturos	Lab.	Practical/		
Торк	liours	Lectures		small groups		
Introduction to Cell and Molecular Biology.						
Cell Structure and Function.						
.Biomolecules						
Enzymes and Metabolism.						
DNA Replication and Repair.						
Transcription and RNA Processing.						
Translation and Protein Folding.						
Genetic Variation and Mutation.						
Recombinant DNA Technology.						
Genetic Engineering.						
Genomics and Proteomics.						
Gene Expression and Regulation.						
Cellular signaling and its regulation.						
Signal Transduction Pathways.						
Genomics and Proteomics.						
.Gene Expression and Regulation						
Cellular Signaling and its Regulation.						
Signal Transduction Pathways.						
Intracellular Vesicular Transport.						
Cell Cycle and Division.						
Apoptosis and Cell Death.						
Cancer Biology.						
Stem Cells and Regeneration.						
Total						

:Learning Methods .5

a. Lectures

b. Tutorial

5 Page 3 of

c. Lab.

6. Assessment methods:

	Assessment Type	Date	%
	First assessment	16 th week of first term.	<mark>30</mark>
	Lab. Exam	Before the final examination	<mark>20</mark>
	Final examination (Written,)	At the end of term	<mark>50</mark>
	100		

7. List of References:

Title	Publisher	сору	authors	Available
				place
Molecular biology	Garland	6th ed	Alberts, B., Johnson, A.,	
of the cell	Science.		Lewis, J., Raff, M., Roberts,	
			K., & Walter, P.	
Molecular cell	W. H. Freeman	8th ed.	Lodish, H., Berk, A.,	
biology	and Company.		Zipursky, S. L., Matsudaira,	
			P., Baltimore, D., & Darnell,	
			J.	

8. Facilities Required for Teaching and Learning:

	Facilities Required	Notes
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Course Coordinators

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Date:...../...../....../

	a. Knowledge and	الاسبوع		
d. General&	c. Professional &	b. Intellectual skills	Understanding	الدراسد
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5 Page 4 of

Head of Department

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The name of university: Programme name: The course name: Course code: Academic year / level: University of Benghazi Molecular diagnostics Molecular Biology Techniques MOLD-204 2023-2024. Second year

6 Page 1 of
1. Basic information:

the course name:	Molecular Biology Techniques
Course coordinator	
Programme (s) on which the course is given:	Molecular diagnostics
Teaching hours	3 hours/week
Language	English
Academic year / level:	2023-2024. Second year
Course approval date	

1.1 hours per week

Lecture: (1 hours/week)

Practical: (2 hours/week) Training: (-) Total: (hours 96)

2. Aims of Course:

- a. To provide students with theoretical and practical knowledge of molecular biology techniques used in research and clinical settings.
- b. To covers essential molecular biology techniques, including DNA extraction, PCR, cloning, gene expression analysis, and DNA sequencing.
- c. To develop students' practical skills in molecular biology and prepare them for research and clinical careers.

3. Intended Learning Outcomes of Course (ILOs):

a- Knowledge and Understanding:

By the end of the course, student should be able to :

- a1. Understand the fundamental concepts and principles of molecular biology.
- a2. Understand the theoretical basis of essential molecular biology techniques.
- a3. Understand the role of molecular biology techniques in research and clinical settings.

b- Intellectual Skills:

By the end of the course, student should be able to :

- b1. Analyze scientific literature related to molecular biology techniques.
- b2. Design experiments and optimize protocols for molecular biology techniques.
- b3. Interpret data generated from molecular biology experiments.

c- Professional and Practical Skills:

By the end of the course, student should be able to :

- c1. Demonstrate practical skills in molecular biology techniques.
- c2. Perform experiments and record data accurately and precisely.
- c3. Work collaboratively in a laboratory setting.

d- General Skills:

By the end of the course, student should be able to:

6 Page 2 of

- d1. Improve communicate scientific ideas and results effectively.
- d2. Enhance problem-solving skills .
- d3. Develop time-management skills.

4- Course Contents:

Tonic	Hours	Locturos	Lab.	Practical/
Торк	liours	Lectures		small groups
Introduction to molecular biology techniques				
DNA Extraction				
Polymerase Chain Reaction (PCR)				
Gel Electrophoresis				
Cloning				
Gene Expression Analysis				
Western Blotting				
Enzyme-Linked Immunosorbent Assay				
(ELISA)				
DNA Sequencing				
Restriction Enzyme Digestion				
Site-Directed Mutagenesis				
RNA Extraction and Analysis				
Real-Time PCR				
Microarray Analysis				
Epigenetic Modifications:				
:Recombinant Protein Production				
Genome Editing				
RNA Interference (RNAi):				
Stem Cell Culture and Differentiation:				
Total				

5. Learning Methods:

- a. Lectures
- b. Tutorial
- c. Lab.

6. Assessment methods:

	Assessment Type	Date	%
1.	First assessment	16 th week of first term.	30
2.	Lab. Exam	Before the final examination	20
3.	Final examination (Written,)	At the end of term	50
	100		

6 Page 3 of

7. List of References:

Title	Publisher	сору	authors	Available place
Molecular Biology		4th Edition	Susan Carson,	
Techniques: A			Heather Miller,	
Classroom			and D. Scott	
Laboratory Manual,			Witherow	
Molecular Biology:		3rd Edition	Michael M.	
Principles and			Cox, Jennifer	
Practice			Doudna, and	
			Michael	
			O'Donnell	

8. Facilities Required for Teaching and Learning:

	Facilities Required	Notes
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Course Coordinators

Head of Department

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The name of university: Program name The course name: Course code: Academic year / level:

University of Benghazi Molecular diagnostics Principles of Human Genetics MOLD-205 2023-2024. Second year

6 Page 1 of

1- Basic information:

the course name:	Principles of Human Genetics
Course coordinator	
Program (s) on which the course is given:	Molecular diagnostics
Teaching hours	4 hours/week
Language	English
Academic year / level:	2023-2024. Second year
Course approval date	

1.1 hours per week

Lecture: (2 hours/week)	Practical: (2 hours/week)	Total: (hours 96)
	Training: (-)	

2- Aims of Course:

- a. The Principles of Human Genetics undergraduate course in Biomedical Sciences aims to provide students with a comprehensive understanding of the fundamental principles of human genetics and their applications in various fields of medicine and healthcare.
- b. The course will cover the basic concepts of genetics, including inheritance patterns, molecular genetics, gene expression, and genomics, and how these principles apply to human health and disease.
- c. The course will also cover ethical, legal, and social issues related to human genetics and genomics.

3- Intended Learning Outcomes of Course (ILOs):

a- Knowledge and Understanding:

After completing this course, students should be able to demonstrate:

- 1- Demonstrate a solid understanding of the fundamental principles of human genetics
- 2- Discuses genetic data using molecular genetic techniques
- 3- Underline principles to the diagnosis and treatment of human diseases
- 4- Understand the ethical, legal, and social issues related to human genetics and genomics

b- Intellectual Skills:

By the end of the course, student should be able to :

- 1- Analyze genetic data using molecular genetic techniques.
- 2- Apply genetic principles to the diagnosis and treatment of human diseases.
- 3- Critically evaluating research studies in human genetics.
- 4- Synthesizing and interpreting complex genetic information.
- 5- Apply hypotheses and designing experiments in genetics research.

c- Professional and Practical Skills:

6 Page 2 of

By the end of the course, student should be able to:

- 1. Genetic counseling and communication skills.
- 2. Designing and implementing genetic tests.
- 3. Interpreting and reporting genetic test results.
- 4. Performing molecular genetic techniques.
- 5. Analyzing and interpreting genetic data using bioinformatics tools.

d- General Skills:

By the end of the course, student should be able to:

- 1- Enhance communication skills.
- 2- Develop teamwork skills.
- 3- Improve time management and organization skills.
- 4- Develop critical thinking, problem-solving, and analytical skills
- 5- Develop professional and practical skills necessary for a career in the biomedical sciences

4- Course Contents:

Торіс	Hours	Lectures	Lab.	Practical/ small groups
Introduction to Human Genetics.				
History of Human Genetics.				
.Structure and Function of DNA				
Chromosome Structure and Function.				
Mendelian Genetics.				
Pedigree Analysis.				
Single Gene Disorders.				
Chromosomal Disorders.				
Carrier Testing.				
Molecular Genetics.				
Gene Expression.				
Gene Regulation.				
Epigenetics.				
Genomics.				
Human Genome Project.				
.Genetic Discrimination				
Personalized Medicine.				
Precision Medicine.				
Pharmacogenetics.				
Genetic Diversity.				
Gene-Environment Interactions.				
Total				

- a. Lectures
- b. Tutorial
- c. Lab.

6. Assessment methods:

Assessment Type	Date	%			
First assessment	16 th week of first term.	<mark>30</mark>			
Lab. Exam	Before the final examination	<mark>20</mark>			
Final examination (Written,)	At the end of term	<mark>50</mark>			
Total					

7. List of References:

Title	Publisher	сору	authors	Available
				place
Essentials of	Pearson		Klug, W. S., Cummings, M.	
Genetics.	Education, Inc.		R., Spencer, C. A., Palladino,	
			M. A.	
Genes IX.	Jones &		Lewin, B.	
	Bartlett			
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8. Facilities Required for Teaching and Learning:

	Facilities Required	Notes
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Course Coordinators

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Head of Department

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The name of university: Program name The course name: Course code: Academic year / level: University of Benghazi Molecular diagnostics Medical Immunology MLSC-205 2023-2024. Second year

6 Page 1 of

1- Basic information:

the course name:	Medical Immunology
Course coordinator	
Program (s) on which the course is given:	Molecular diagnostics
Teaching hours	4 hours/week
Language	English
Academic year / level:	2023-2024. Second year
Course approval date	

1.1 hours per week

Lecture: (2 hours/ week)

Practical: (2 hours/ week) Training: (-) Total: (hours 96)

1- Aims of Course:

- a) The course aims to provide students with a comprehensive understanding of the immune system, its mechanisms, functions, and clinical applications.
- b) The course intends to equip students with knowledge and skills to enable them to pursue careers in biomedical sciences, immunology research, and clinical practice.
- c) The course aims to develop students' ability to think critically and analytically, communicate effectively, and work collaboratively in a team.

2- Intended Learning Outcomes of Course (ILOs):

a- Knowledge and Understanding:

After completing this course, students should be able to demonstrate:

- 1. Describe the components, organization, and functions of the immune system.
- 2. Understand the molecular basis of immune recognition and response.
- 3. Discuss the role of immune system in health and disease.
- 4. Describe the genetic, ontogenic, and clinical implications of immune system function.

b- Intellectual Skills:

By the end of the course, student should be able to:

- 1. Analyze and interpret immunological data and literature.
- 2. Evaluate the contribution of immune system in disease pathogenesis.
- 3. Design and execute immunological experiments.

c- Professional and Practical Skills:

By the end of the course, student should be able to:

- 1. Illustrate immunological concepts and findings.
- 2. Practice laboratory skills and techniques in immunological research.
- 3. Explain ethical and professional behavior in biomedical sciences.

d- General Skills:

By the end of the course, student should be able to:

- 1. Develop effective written and oral communication skills.
- 2. Enhance time-management and organization skills.
- 3. Develop computer literacy and data analysis skills.
- 4. Develop leadership and team management skills.
- 5. Develop critical thinking and problem-solving skills.

4- Course Contents:

Торіс	Hours	Lectures	Lab	Practical/ small
				groups
Introduction				
Leucocytes and Lymphoid Tissues: The				
Framework of the Immune System				
Major Histocompatibility Complex				
The Adaptive Immune Response: Antigens,				
Lymphocytes and Accessory Cells				
Immunoglobulins: Structure and Diversity				
Immunoglobulins: Metabolism and Biological				
Properties				
Genetics of Immunoglobulins: Ontogenic,				
Biological and Clinical Implications				
Antigen-Antibody Reactions				
The Complement System in Health and Disease				
Monocyte and Lymphocyte Membrane Markers:				
Ontogeny and Clinical Significance				
Cell-Mediated Immunity				
Adaptive Humoral Immunity and				
Immunoprophylaxis				
Phagocytic Cells and their Functions				
Anti-Infectious Innate and Adaptive Immune				
Responses				
Diagnostic Applications of Immunology				
Tolerance and Autoimmunity				
Organ-Specific Autoimmune Diseases				
Systemic Lupus Erythematosus				
Rheumatoid Arthritis				
Overview of Hypersensitivity				
IgE-mediated (Immediate) Hypersensitivity				
Immunohematology				
The Pathogenic Role				
of Antigen-Antibody				

6 Page 3 of

			Lab	Practical/
Торіс	Hours	Lectures		small
				groups
Complexes				
Immune System Modulators				
Transplantation Immunology				
Tumor Immunology				
Lymphocyte and Plasma Cell malignancies				
Diagnosis of Immune Deficiency Diseases				
Primary Immunodeficiency Diseases				
AIDS and Other Acquired Immunodeficiencies				
Total.				

5. Learning Methods:

- a. Lectures
- b. Tutorial
- c. Lab.

6. Assessment methods:

Assessment Type	Date	%
First assessment	16 th week of first term.	<mark>30</mark>
Lab. Exam	Before the final examination	<mark>20</mark>
Final examination (Written,)	At the end of term	50
T.	otal	<mark>100</mark>

7. List of References:

Title	Publisher	сору	authors	Available
				place
Medical Immunology.	CRC Press.	7th Ed.	Virella, G	

8. Facilities Required for Teaching and Learning:

	Facilities Required	Notes
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Course Coordinators

Head of Department

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The name of university: Programme name: The course name: Course code: Academic year / level: University of Benghazi Molecular diagnostics Bioinformatics and Genomics MOLD-301 2023-2024. Third year

5 Page 1 of

1. Basic information:

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the course name:	Bioinformatics and Genomics
Course coordinator	
Programme (s) on which the course is given:	Molecular diagnostics
Teaching hours	5 hours/week
Language	English
Academic year / level:	2023-2024. Third year
Course approval date	

1.1 hours per week

Lecture: (3 hours/week)

Practical: (2 hours/week) Training: (-) Total: (hours 160)

2. Aims of Course:

- a. To provide students with a deep understanding of the principles and practices of bioinformatics, genomics, and computational biology.
- b. To provide students with a solid foundation in the underlying concepts of molecular biology, genetics, and computer science, enabling them to apply this knowledge in the analysis of complex biological data.
- c. To gain practical experience in the use of software tools and databases, and develop professional and transferable skills in data analysis, interpretation, and communication.

3. Intended Learning Outcomes of Course (ILOs):

a- Knowledge and Understanding:

By the end of the course, student should be able to :

1. Understand deeply the molecular basis of life and the relationship between genomics, proteomics, and transcriptomics.

2. Outline the basic principles and methods of sequence alignment, gene prediction, and sequence analysis.

3.Decribe usage of computational techniques to analyze and interpret large datasets.

4. Identify and use databases, software tools, and other resources for the analysis of biological data.

b- Intellectual Skills:

By the end of the course, student should be able to :

1. Develop critical thinking skills for the analysis of complex biological data.

2. Apply computational and statistical methods to interpret biological data.

3. Analyze complex genetic and genomic data and draw meaningful conclusions.

4. Develop the ability to identify and address biological questions using bioinformatics and genomics approaches.

5 Page 2 of

c- Professional and Practical Skills:

By the end of the course, student should be able to :

- 1. Interpret bioinformatics tools and databases.
- 2. Analysis and interpretation of biological data.
- 3. Summarize scientific findings.
- 4. Operate team-based research and collaboration.

d- General Skills:

By the end of the course, student should be able to:

- 1. Develop skills in problem-solving, critical thinking, and data analysis.
- 2. Improve scientific writing and communication skills.
- 3. Enhance time management.

4- Course Contents:

Tonic	Hours	Locturos	Lab.	Practical/
Topic	mours	Lectures		small groups
1: Introduction to Bioinformatics and				
Genomics.				
2: Molecular Biology and Genetics				
Sequence Alignment .3				
4: Sequence Analysis				
5: Gene Prediction				
6: Genome Assembly				
7: Comparative Genomics				
8: Transcriptomics				
9: Proteomics				
10: Metabolomics				
11: Structural Bioinformatics				
12: Systems Biology				
13: Genomic Variation				
14: Epigenomics				
15: Functional Genomics				
Next-Generation Sequencing :16				
17: High-Performance Computing				
18: Machine Learning				
19: Network Analysis				
20: Pathway Analysis				
21: Data Visualization				
22: Bioinformatics Databases.				
23: Personalized Genomics				
24: Current Topics in Bioinformatics and				
Genomics.				
Total				

5 Page 3 of

:Learning Methods .5

- a. Lectures
- b. Tutorial
- c. Lab.

6. Assessment methods:

	Assessment Type	Date	%				
1.	First assessment	16 th week of first term.	30				
2.	Lab. Exam	Before the final examination	20				
3.	Final examination (Written,)	At the end of term	50				
	Total						

7. List of References:

Title	Publisher	сору	authors	Available place
Bioinformatics and	John Wiley &	3^{rd} .ed.	Pevsner, J	
functional genomics	Sons			
Introduction to	Oxford	4th ed	Lesk, A. M.	
bioinformatics	University			
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8. Facilities Required for Teaching and Learning:

	Facilities Required	Notes
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Course Coordinators

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Head of Department

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The name of university: Programme name: The course name: Course code: Academic year / level: University of Benghazi Molecular diagnostics Research Methodology and Data Analysis BMSC-301 2023-2024. Third year

7 Page 1 of

1. Basic information:

the course name:	Research Methodology and Data Analysis
Course coordinator	
Programme (s) on which the course is given:	Molecular diagnostics
Teaching hours	5 hours/week
Language	English
Academic year / level:	2023-2024. Third year
Course approval date	

1.1 hours per week

Lecture: (3 hours/week)

Practical: (2 hours/week) Training: (-) Total: (hours 160)

2. Aims of Course:

- **a.** To provide students with an extensive comprehension of research methodologies and data analysis techniques applicable to various disciplines.
- **b.** to equip students with the skills required to develop research proposals, conduct research, gather data, perform data analysis, and interpret results.
- c. To `emphasizes improving students' critical thinking and problem-solving abilities.
- **d.** To impart knowledge about research methodologies and data analysis techniques applicable to medical research, thereby enabling students to develop research proposals, conduct research, collect data, analyze data, and interpret results in the medical field.

3. Intended Learning Outcomes of Course (ILOs):

a- Knowledge and Understanding:

By the end of the course, student should be able to :

1. Understand the basic principles of research methodology in biomedical fields,

including study design, sampling, data collection, and analysis

2. Outline of the concepts and theories related to research methodology and data analysis in biomedical sciences

3. Demonstrate intellectual skills in analyzing and evaluating research literature,

formulating research questions and hypotheses, and selecting appropriate research methodologies

4. Develop professional and practical skills in designing and conducting research projects, collecting and analyzing data, and presenting research findings

5. Understand the principles of scientific writing and research communication in biomedical fields

b- Intellectual Skills:

By the end of the course, student should be able to :

b1. Develop critical thinking and problem-solving skills through analyzing and interpreting research data

7 Page 2 of

b2. Develop the ability to formulate research questions, hypotheses, and objectives

b3. Develop the ability to design and implement research studies in biomedical fields

b4. Develop the ability to evaluate the strengths and limitations of different research methodologies in biomedical fields

c- Professional and Practical Skills:

By the end of the course, student should be able to :

c1. Develop the skills to effectively communicate research findings to a range of audiences

c2. Develop the skills to write scientific research proposals and manuscripts in biomedical fields

c3. Develop the skills to analyze and interpret research data using statistical software and other tools

c4. Develop the skills to manage and organize research data in biomedical fields

d- General Skills:

By the end of the course, student should be able to:

d1. Develop general transferable skills in critical thinking, problem-solving,

communication, and teamwork

d2. Improve teamwork and collaboration skills through group projects and presentations

d3. Enhance time management and organizational skills through planning and executing research projects

d4. Develop problem-solving and analytical skills that can be applied to a range of fields

			Lab.	Practical/
Торіс	Hours	Lectures		small
				groups
1. Introduction to Research				
2. Research Methodology: An Introduction				
3. Reviewing the Literature				
4. Writing Protocol				
5. Formulating a Research Problem and				
Research Question				
6. Ethics in Research				
7. Introduction to Epidemiology				
8. Measurement Tools in Epidemiology				
9. Descriptive Studies				
10. Case-Control Studies				
11. Cohort Studies				
12. Questionnaire				

4- Course Contents:

7 Page **3** of

— •	H	Testeres	Lab.	Practical/
Горіс	Hours	Lectures		small grouns
Design				Stoups
13. Medical Reading and Writing				
14. Structure of a Medical Research Paper				
15. Screening				
16. Association and Causation				
17. Basic Concepts and Definitions in Statistics				
18. Selecting a Method of Data Collection				
19. Descriptive Statistics				
20. Inferential Statistics and Significant Test				
21. The Use of Excel and SPSS				
22. Research Proposal Projects				
Total				

:Learning Methods .5

- a. Lectures
- b. Tutorial
- c. Lab.

6. Assessment methods:

	Assessment Type	Date	%			
1.	First assessment	16 th week of first term.	30			
2.	Lab. Exam	Before the final examination	20			
3.	Final examination (Written,)	At the end of term	50			
Total 100						

7. List of References:								
	Publisher	сору	authors	Available place				

7 Page 4 of

Title			
Introduction to	How To Books	Dawson, C.	
research methods: A	Ltd.		
practical guide for			
anyone undertaking			
a research project.			
Research methods in	Routledge.	Gliner, J. A.,	
applied settings: An		Morgan, G. A.,	
integrated approach		& Leech, N. L.	
to design and		,	
analysis.		 	
Research	Sage	Kumar, R.	
Methodology: A	Publications		
Step-by-Step Guide			
for Beginners.			

8. Facilities Required for Teaching and Learning:

	Facilities Required	Notes
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Course Coordinators

Head of Department

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7 Page 7 of





The name of university: Programme name: The course name: Course code: Academic year / level: University of Benghazi Molecular diagnostics Clinical Molecular Diagnostic Techniques MOLD-303 2023-2024. Third year

5 Page 1 of

1. Basic information:

the course name:	Clinical Molecular Diagnostic Techniques
Course coordinator	
Programme (s) on which the course is given:	Molecular diagnostics
Teaching hours	3 hours/week
Language	English
Academic year / level:	2023-2024. Third year
Course approval date	

1.1 hours per week

Lecture: (1 hours/week)

Practical: (2 hours/week) Training: (-) Total: (hours 96)

2. Aims of Course:

- **a.** To provide students with a comprehensive understanding of the principles and applications of molecular diagnostics in the diagnosis and management of diseases.
- **b.** To provide students with hands-on experience in laboratory techniques for the detection of genetic mutations, infectious agents, and other disease markers. By the end of the course,
- **c.** to demonstrate a detailed knowledge of the principles and applications of molecular diagnostics, as well as an ability to apply this knowledge in laboratory settings

3. Intended Learning Outcomes of Course (ILOs):

a- Knowledge and Understanding:

By the end of the course, student should be able to :

1. Outline the principles and applications of molecular diagnostics.

2. Understand the molecular basis of genetic mutations, infectious agents, and other disease markers.

3. Describe and interpret molecular diagnostic data to inform the diagnosis and management of disease.

b- Intellectual Skills:

By the end of the course, student should be able to :

- 1. Evaluate the scientific literature on molecular diagnostics and its applications.
- 2. Apply theoretical knowledge to practical problem-solving scenarios.
- 3. Formulate and test hypotheses based on molecular diagnostic data.
- 4. Analyze and interpret complex molecular diagnostic datasets.

c- Professional and Practical Skills:

By the end of the course, student should be able to :

1. Practice molecular diagnostic techniques in a laboratory setting, including nucleic acid extraction, PCR, sequencing, and other techniques.

- 2. Interpret molecular diagnostic data to inform clinical decision making.
- 3. Rewrite scientific concepts and research findings effectively in written and oral formats.

5 Page 2 of

4.Practice effectively in a laboratory environment, including following safety protocols and good laboratory practices.

d- General Skills:

By the end of the course, student should be able to:

- 1. Develop effective time-management and organizational skills.
- 2. Improve effective problem-solving and critical thinking skills.
- 3. Develop team-working and collaboration skills.

4- Course Contents:

Tonio	Houng	Lasturas	Lab.	Practical/
Торіс	nours	Lectures		small groups
Introduction to Molecular Diagnostics.				
Nucleic Acid Extraction.				
.PCR and qPCR				
Sequencing.				
Microarrays.				
Next-Generation Sequencing.				
Digital PCR .				
CRISPR/Cas9.				
Biomarkers.				
Gene Expression Profiling.				
Epigenetics.				
Infectious Disease Diagnostics.				
Cancer Diagnostics.				
Cardiovascular Disease Diagnostics.				
Neurological Disease Diagnostics.				
.Respiratory Disease Diagnostics				
Renal Disease Diagnostics.				
Digestive Disease Diagnostics.				
Reproductive Health Diagnostics.				
Inherited Genetic Disorder Diagnostics.				
Emerging Technologies in Molecular				
Diagnostics.				
Total				

:Learning Methods .5

- a. Lectures
- b. Tutorial
- c. Lab.

5 Page 3 of

6. Assessment methods:

	Assessment Type	Date	%
1.	First assessment	16 th week of first term.	30
2.	Lab. Exam	Before the final examination	20
3.	Final examination (Written,)	At the end of term	50
	100		

7. List of References:

Title	Publisher	сору	authors	Available place
Molecular	F.A. Davis		Buckingham,	
Diagnostics:	Company.		L., & Flaws,	
Fundamentals,			M. L.	
Methods, and				
Clinical				
Applications.				
Clinical Molecular	Academic		Coleman, W.	
Diagnostics: The	Press		B., &	
Essentials.			Tsongalis, G.	
			J.	

8. Facilities Required for Teaching and Learning:

	Facilities Required	Notes
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Course Coordinators

Head of Department

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The name of university: Programme name: The course name: Course code: Academic year / level:

University of Benghazi Molecular diagnostics Medical Genetics MOLD-304 2023-2024. Third year

6 Page 1 of

1. Basic information:

the course name:	Medical Genetics
Course coordinator	
Programme (s) on which the course is given:	Molecular diagnostics
Teaching hours	4 hours/week
Language	English
Academic year / level:	2023-2024. Third year
Course approval date	

1.1 hours per week

Lecture: (2 hours/week)

Practical: (2 hours/week) Training: (-) Total: (hours 128)

2. Aims of Course:

- **a.** To provide students with an in-depth understanding of the principles and applications of genetics in medicine, with a focus on molecular, cytogenetic, immunogenetic, population, and mathematical genetics.
- **b.** to provide students with an understanding of the history and impact of genetics in medicine, DNA technology and its applications, methods of chromosome analysis, immune genetics, and genetic disorders.
- **c.** to provide students with knowledge about the ethical, legal, and social issues in genetics, genetic counseling, and genetic testing, as well as the application of genetics in society.
- d.

3. Intended Learning Outcomes of Course (ILOs):

a- Knowledge and Understanding:

By the end of the course, student should be able to :

- 1. Recognize the history and impact of genetics in medicine.
- 2. Define DNA as the basis of inheritance and the structure and function of genes.

3. Outline of the types of genetic disorders, chromosome abnormalities, and patterns of inheritance.

4. Describe DNA technology and its applications, including techniques of DNA analysis, DNA sequencing, and the application of DNA sequence polymorphisms.

5. Understand immune genetics, including innate and specific acquired immunity, blood groups, HLA, and immunology of transplantation.

6. Underline population and mathematical genetics, including the Hardy-Weinberg principle, factors that alter gene frequency, consanguinity, segregation analysis, genetic linkage, and risk calculation.

7. Understand genetic disorders, including hemoglobinopathies, disorders of coagulation and bleeding, and biochemical genetics.

b- Intellectual Skills:

By the end of the course, student should be able to :

1. Evaluate critically scientific literature related to genetics in medicine.

6 Page **2** of
2. Analyze data obtained from DNA sequencing and DNA analysis techniques.

3. Perform segregation analysis and calculate risks of inheritance.

c- Professional and Practical Skills:

By the end of the course, student should be able to :

1. Practice genetic counseling and genetic testing, including preimplantation genetic diagnosis,

prenatal diagnosis and screening, carrier testing, fetal DNA analysis, and preconception counseling.

2. Operate newborn screening and pharmacogenetic testing.

3. Demonstrate the ethical, legal, and social issues in genetics and apply this knowledge in clinical practice.

4. Discuss the importance of genetics in medicine with healthcare professionals, patients, and the general public.

d- General Skills:

By the end of the course, student should be able to:

1. Develop a teamwork silks with other healthcare professionals.

3. Enhance the ability to adapt to new technologies and techniques in the field of genetics.

4- Course Contents:

Tonia	Hours	Looturos	Lab.	Practical/
Торіс	nours	Lectures		small groups
A. Principles of Human Genetics (review)				
Basic Cell Biology: Structure and Function				
of genes				
Chromosomes and Cell Division				
Patterns of Inheritance				
B. Molecular Genetics				
Mapping and Identifying Genes for				
Monogenic Disorders				
C. Cytogenetics				
D. Immunogenetics				
E. Population And Mathematical Genetics				
Genetics In Medicine				
2 Disorders of coagulation and bleeding				
3 Biochemical Genetics				
G. Genetics In Society				
Diagnosis and Screening, Carrier testing,				
Fetal DNA analysis, Preconception				
counseling				
Total				

5. Learning Methods:

- a. Lectures
- b. Tutorial

6 Page 3 of

c. Lab.

6. Assessment methods:

	Assessment Type	Date	%
1.	First assessment	16 th week of first term.	30
2.	Lab. Exam	Before the final examination	20
3.	Final examination (Written,)	At the end of term	50
	Т	otal	100

7. List of References:

Title	Publisher	сору	authors	Available place
Emery's Elements of	Elsevier	16th edition.	Peter	
Medical Genetics			Turnpenny,	
			Sian Ellard,	
			and Ruth	
			Lovering	
Medical Genetics at	Wiley-	4th edition.	Dorian J.	
a Glance	Blackwell		Pritchard and	
			Bruce R. Korf.	

8. Facilities Required for Teaching and Learning:

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Course Coordinators

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Head of Department

Date:...../...../....../

6 Page 4 of

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6 Page 6 of





The name of university: Programme name: The course name: Course code: Academic year / level: University of Benghazi Molecular diagnostics Histotechnology CYTO-305 2023-2024. Third year

6 Page 1 of

1. Basic information:

the course name:	Histotechnology
Course coordinator	
Programme (s) on which the course is given:	Molecular diagnostics
Teaching hours	4 hours/week
Language	English
Academic year / level:	2023-2024. Third year
Course approval date	

1.1 hours per week

Lecture: (2 hours/week)

Practical: (2 hours/week) Training: (-) Total: (hours 128)

2. Aims of Course:

- **a.** To introduce students to the fundamental principles and techniques used in the preparation, processing, and staining of biological tissues.
- **b.** to provide students with a strong foundation in the theoretical and practical aspects of histotechnology.
- **c.** to develop their professional and transferable skills.

3. Intended Learning Outcomes of Course (ILOs):

a- Knowledge and Understanding:

By the end of the course, student should be able to :

1. Outline the principles of microtechnique and the role of histology in biomedical sciences

2. Describe different types of fixatives and their effects on tissue structure and antigenicity

3. Underline various methods used for decalcification and other treatments for hard tissues.

- 4. Understand steps involved in tissue processing and mounting
- 5. Define types of dyes used in histotechnology and their staining mechanisms
- 6. Identify staining techniques used for tissues, blood, and other cell suspensions

7. Describe methods used for connective tissues, nucleic acids, organic functional groups, carbohydrates, lipids, pigments, and inorganic ions

8. Outline principles of enzyme histochemistry and the different classes of enzymes

9. Identify the methods used for soluble organic compounds and metal reduction and precipitation

10. State the principles of immunohistochemistry and the different types of antibodies

b- Intellectual Skills:

By the end of the course, student should be able to :

1. Analyze and interpret histological images and data

6 Page **2** of

- 2. Apply critical thinking to solve problems in histotechnology
- 3. Evaluate the quality of tissue samples and staining protocols

c- Professional and Practical Skills:

By the end of the course, student should be able to :

- 1. Perform tissue processing, embedding, and sectioning
- 2. Prepare and use staining solutions for histological samples
- 3. Perform enzyme histochemistry and immunohistochemistry techniques

d- General Skills:

By the end of the course, student should be able to:

- 1. Improve teamwork skills
- 2. Enhance communicate scientific ideas effectively in written and oral formats.
- 3. Develop effective time management and organizational skills.

4- Course Contents:

Tonic	Hours	Locturos	Lab.	Practical/
Торк	IIIUIIS	Lectures		small groups
Introduction to micro-technique				
Fixation				
Decalcification and other treatments for hard				
tissues				
Processing and mounting				
Dyes				
Staining with dyes in one or two colours				
Staining blood and other cell suspensions				
Methods for connective tissue				
Methods for nucleic acids				
Organic functional groups and protein				
histochemistry				
Carbohydrate histochemistry				
Lipids				
Methods for pigments and inorganic ions				
Enzyme histochemistry: general				
considerations				
Hydrolytic enzymes				
Oxidoreductases				
Methods for soluble organic compounds of				
low molecular weight				
Metal reduction and precipitation methods				
Immunohistochemistry				
Special techniques				
Total				

6 Page 3 of

:Learning Methods .5

- a. Lectures
- b. Tutorial
- c. Lab.

6. Assessment methods:

	Assessment Type	Date	%						
1.	First assessment	16 th week of first term.	30						
2.	Lab. Exam	Before the final examination	20						
3.	Final examination (Written,)	At the end of term	50						
	Total								

7. List of References:

Title	Publisher	сору	authors	Available place
Theory and Practice	Churchill	6th edition	Bancroft, J.D.	
of Histological	Livingstone,		and Gamble,	
Techniques	Elsevier		M.	
Histological and	Scion	5th edition.	Kiernan, J.A.	
Histochemical	Publishing Ltd,			
Methods: Theory				
and Practice.				

8. Facilities Required for Teaching and Learning:

	Facilities Required	Notes
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Course Coordinators

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Head of Department

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6 Page 6 of





The name of university: Programme name: The course name: Course code: Academic year / level:

University of Benghazi Molecular diagnostics Molecular Basis of Disease MOLD-305 2023-2024. Third year

5 Page 1 of

1. Basic information:

the course name:	Molecular Basis of Disease
Course coordinator	
Programme (s) on which the course is given:	Molecular diagnostics
Teaching hours	5 hours/week
Language	English
Academic year / level:	2023-2024. Third year
Course approval date	

1.1 hours per week

Lecture: (3 hours/week)

Practical: (2 hours/week) Training: (-) Total: (hours 160)

2. Aims of Course:

- **a.** To provide students with an in-depth understanding of the molecular mechanisms that underlie human disease.
- **b.** to introduce students to a wide range of molecular and cellular processes that contribute to the development and progression of disease.
- **c.** students should be able to analyze and interpret the molecular mechanisms underlying a wide range of diseases.

3. Intended Learning Outcomes of Course (ILOs):

a- Knowledge and Understanding:

By the end of the course, student should be able to :

1. Understand the molecular basis of human disease.

2. Outline the key molecular and cellular processes that contribute to the development and progression of disease.

3. Describe the genetic and environmental factors that contribute to disease susceptibility.

4. Underline the molecular mechanisms underlying the pathogenesis of a range of human diseases.

b- Intellectual Skills:

By the end of the course, student should be able to :

- 1. Evaluate critically the scientific literature on the molecular basis of disease.
- 2. Relate theoretical knowledge to practical problem-solving scenarios.
- 3. Formulate and test hypotheses based on molecular data.
- 4. Analyze and interpret complex datasets from a range of molecular techniques.

c- Professional and Practical Skills:

By the end of the course, student should be able to :

1. Interpret scientific concepts and research findings effectively in written and oral formats.

2. Practice in a laboratory environment, including handling of biological materials and molecular techniques.

5 Page 2 of

3. Demonstrate the usage of common laboratory techniques in molecular biology and disease research.

d- General Skills:

By the end of the course, student should be able to:

- 1. Improve independent learning skills.
- 2. Develop effective time-management and organizational skills.
- 3. Develop effective problem-solving and critical thinking abilities.
- 4. Enhance written and oral communication skills.
- 5. Develop team-working and collaboration skills.

4- Course Contents:

Tonic	Hours	Locturos	Lab.	Practical/
Topic	liours	Lectures		small groups
1: Introduction to Bioinformatics and				
Genomics.				
Introduction to Molecular Basis of Disease.				
.DNA Replication, Repair and Mutations				
Cell Cycle and Cancer.				
Apoptosis and Cell Death.				
Protein Folding and Misfolding.				
Prion Diseases.				
Protein Trafficking and Secretion.				
Metabolism and Disease.				
Diabetes				
Immune Response and Inflammation.				
Systemic Inflammation and Cancer				
Infectious Diseases.				
.Neurodegenerative				
Cardiovascular Diseases.				
Respiratory Diseases.				
Digestive Diseases .				
Endocrine Diseases.				
Renal Diseases.				
Musculoskeletal Diseases.				
Reproductive Health and Disease.				
Genetic Diseases.				
Total				

:Learning Methods .5

a. Lectures

5 Page 3 of

- b. Tutorial
- c. Lab.

6. Assessment methods:

	Assessment Type	Date	%			
1.	First assessment	16 th week of first term.	30			
2.	Lab. Exam	Before the final examination	20			
3.	Final examination (Written,)	At the end of term	50			
	Total					

7. List of References:

Title	Publisher	сору	authors	Available place
Molecular biology	Garland	6th ed.	Alberts, B.,	
of the cell	Science.		Johnson, A.,	
			Lewis, J.,	
			Morgan, D.,	
			Raff, M.,	
			Roberts, K., &	
			Walter, P.	
Robbins and Cotran	Elsevier.	10th ed	Kumar, V.,	
pathologic basis of			Abbas, A. K.,	
disease			& Aster, J. C.	

8. Facilities Required for Teaching and Learning:

	Facilities Required	Notes
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Course Coordinators

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	a. Knowledge and	الاسبوع		
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Head of Department

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5 Page 5 of





The name of university: Programme name: The course name: Course code: Academic year / level: University of Benghazi Molecular diagnostics Molecular Immunology MOLD-307 2023-2024. Third year

6 Page 1 of

1. Basic information:

the course name:	Molecular Immunology
Course coordinator	
Programme (s) on which the course is given:	Molecular diagnostics
Teaching hours	5 hours/week
Language	English
Academic year / level:	2023-2024. Third year
Course approval date	

1.1 hours per week

Lecture: (3 hours/week)

Practical: (2 hours/week) Training: (-) Total: (hours 160)

2. Aims of Course:

- **a.** To provide students with an in-depth understanding of the molecular and cellular basis of the immune system.
- **b.** To cover a wide range of topics related to immunology, including immune cell development, antigen recognition, signaling pathways, and immune responses to pathogens and tumors.
- c. students with hands-on experience in a variety of immunology techniques.

3. Intended Learning Outcomes of Course (ILOs):

a- Knowledge and Understanding:

By the end of the course, student should be able to :

1. Identify the molecular and cellular basis of the immune system.

2. Understand the mechanisms of immune cell development, antigen recognition, and signaling pathways.

3. Outline the immune responses to pathogens and tumors.

4. Define the principles of immunotherapy and vaccine development.

b- Intellectual Skills:

By the end of the course, student should be able to :

- 1. Evaluate critically scientific literature in the field of immunology.
- 2. Analyze and interpret experimental data in the context of immunology.
- 3. Formulate hypotheses and design experiments to test them.
- 4. Illustrate professional and practical Skills:
- 5. Demonstrate proficiency in laboratory techniques used in the field of immunology.
- 6. Construct scientific ideas and data effectively in both oral and written formats.

c- Professional and Practical Skills:

By the end of the course, student should be able to :

- c1. Practice a range of immunology techniques.
- c2. Operate effectively in a laboratory environment.

6 Page **2** of

c3. Review laboratory reports and scientific papers.

d- General Skills:

By the end of the course, student should be able to:

- 1. Develop critical thinking and problem-solving skills.
- 2. Develop time-management and organization skills.
- 3. Improve written and oral communication skills.

4- Course Contents:

Торіс	Hours	Lectures	Lab.	Practical/ small groups
1. Properties and Overview of Immune Responses				
2. Cells and Tissues of the Immune System				
3. Leukocyte Circulation and Migration Into Tissues				
4. Innate Immunity				
5. Antibodies and Antigens				
6. Antigen Presentation to T Lymphocytes and the Function of Major Histocompatibility				
7. Immune Receptors and Signal Transduction				
8. Lymphocyte Development and Antigen Receptor Gene Rearrangement				
9. Activation of T Lymphocytes				
10. Differentiation and Functions of CD4+ Effector T Cells				
11. Differentiation and Functions of CD8+ Effector T Cells				
12. B Cell Activation and Antibody Production				
13. Effector Mechanisms of Humoral Immunity				
Specialized Immunity at Epithelial .14 Barriers and in Immune Privileged Tissues				
15. Immunologic Tolerance and Autoimmunity				
16. Immunity to Microbes				
17. Transplantation Immunology				
18. Tumor Immunology				
19. Hypersensitivity Disorders				
20. Allergy				
21. Primary and Acquired				
Immunodeficiencies				

:Learning Methods .5

- a. Lectures
- b. Tutorial
- c. Lab.

6. Assessment methods:

	Assessment Type	Date	%			
1.	First assessment	16 th week of first term.	30			
2.	Lab. Exam	Before the final examination	20			
3.	Final examination (Written,)	At the end of term	50			
	Total					

7. List of References:

Title	Publisher	сору	authors	Available place
Immunobiology: the	Garland		Janeway, C.	
immune system in	Science		A., Travers, P.,	
health and disease.			Walport, M., &	
			Shlomchik, M.	
			J.	
Janeway's	Garland		Murphy, K., &	
immunobiology.	Science.		Weaver, C.	

8. Facilities Required for Teaching and Learning:

	Facilities Required	Notes
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Course Coordinators

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Head of Department

Date:...../...../....../

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6 Page 6 of





The name of university: Programme name: The course name: Course code: Academic year / level: University of Benghazi Molecular diagnostics Specimen Procurement and Documentation MLSC-307 2023-2024. Third year

6 Page 1 of

1. Basic information:

the course name:	Specimen Procurement and
	Documentation
Course coordinator	
Programme (s) on which the course is given:	Molecular diagnostics
Teaching hours	3 hours/week
Language	English
Academic year / level:	2023-2024. Third year
Course approval date	

1.1 hours per week

Lecture: (1 hours/week)

Practical: (2 hours/week) Training: (-) Total: (hours 96)

2. Aims of Course:

- **a.** To provide students with the knowledge and practical skills necessary for the collection, handling, and documentation of specimens in a clinical laboratory setting.
- **b.** To cover a range of topics related to the proper handling and processing of biological specimens, including blood, urine, stool, and tissue samples.
- **c.** To prepare students for careers in clinical laboratory science by providing them with a strong foundation in the theory and practice of specimen procurement and documentation.

3. Intended Learning Outcomes of Course (ILOs):

a- Knowledge and Understanding:

By the end of the course, student should be able to :

1. Understand the principles and methods used in the collection, handling, and documentation of biological specimens

2. List the different types of specimens and their unique handling requirements

3. Describe the role of the clinical laboratory in the healthcare system

4. Outline of the ethical and legal considerations surrounding the handling of biological specimens

b- Intellectual Skills:

By the end of the course, student should be able to :

- 1. Analyze and interpret laboratory data
- 2. Apply critical thinking to specimen collection and handling
- 3. Evaluate quality assurance and control techniques in the laboratory

c- Professional and Practical Skills:

By the end of the course, student should be able to :

- 1. Apply safe and accurate measures to collect, handle, and document biological specimens
- Use and maintain laboratory equipment related to specimen procurement and documentation
 Operate effectively with healthcare professionals and laboratory staff

6 Page 2 of

4. Demonstrate proper techniques for specimen procurement and documentation

5. Practice occupational health and safety measures in the laboratory

6. Apply quality assurance and control techniques in the laboratory

d- General Skills:

By the end of the course, student should be able to:

- 1. Develop effective communication skills in a laboratory setting
- 2. Enhance teamwork skills
- 3. Develop time management and organizational skills

4- Course Contents:

Торіс	Hours	Lectures	Lab.	Practical/ small groups
1. Introduction to Specimen Procurement and Documentation				
2. Types of Specimens				
3. Patient Identification and Specimen Labeling				
4. Specimen Collection and Handling				
5. Specimen Transport and Storage				
6. Quality Assurance and Control				
7. Occupational Health and Safety				
8. Legal and Ethical Issues				
9. Point of Care Testing				
10. Clinical Laboratory Workflow				
Laboratory Information Systems				
Hematology				
Venipuncture:				
Capillary Blood Collection:				
Microbiology				
Clinical Chemistry				
Urinalysis				
Immunology and				

6 Page 3 of

Tonia	Hours	Lasturas	Lab.	Practical/
горіс	nours	Lectures		small groups
Serology				
Cytology and Histology				
Specimen Rejection and Unsatisfactory Samples				
Specimen Referral and External Testing				
Newborn Screening				
Specimen Collection in Special Populations				
Molecular Diagnostics				
Hematopoietic Stem Cell Collection and Processing				
Blood Transfusion				
Collection of Specimens for Drug Monitoring and Toxicology				
Specimen Collection and Transport in Public Health				
Quality Improvement in Specimen Procurement and Documentation				
Emerging Technologies in Specimen Procurement and Documentation				
Total				

5. Learning Methods:

- a. Lectures
- b. Tutorial
- c. Lab.

6. Assessment methods:

	Assessment Type	Date	%
1.	First assessment	16 th week of first term.	30
2.	Lab. Exam	Before the final examination	20
3.	Final examination (Written,)	At the end of term	50
	Т	otal	100

7. List of References:

Title	Publisher	Сору	authors	Available place
Microbiology: a	McGraw-Hill		Nester, E. W.,	
human perspective.	Education.		Anderson, D.	
			G., Roberts, C.	
			E., & Nester,	
			M. T.	
Clinical chemistry:	Elsevier.		Kaplan, L. A.,	
theory, analysis,			& Pesce, A. J.	
correlation.				
Clinical Chemistry:	Lippincott		Bishop M,	
Principles,	Williams &		Fody E,	
Techniques,	Wilkins		Schoeff L.	
Correlations.				
Clinical and	Wayne,	5th Ed.		
Laboratory				
Standards Institute				
(CLSI). Collection,				
Transport, and				
Processing of Blood				
Specimens for				
Testing Plasma-				
Based Coagulation				
Assays and				
Molecular				
Hemostasis Assays;				
Approved				
Guideline,				

8. Facilities Required for Teaching and Learning:

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Course Coordinators

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Head of Department

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The name of university: Programme name: The course name: Course code: Academic year / level: University of Benghazi Molecular diagnostics Molecular Oncology MOLD-308 2023-2024. Third year

6 Page 1 of

1. Basic information:

The course name:	Molecular Oncology
Course coordinator	
Programme (s) on which the course is given:	Molecular diagnostics
Teaching hours	4 hours/week
Language	English
Academic year / level:	2023-2024. Third year
Course approval date	

1.1 hours per week

Lecture: (2 hours/week)

Practical: (2 hours/week) Training: (-) Total: (hours 128)

2. Aims of Course:

- **a.** To provide students with a comprehensive understanding of the molecular mechanisms that underlie the development and progression of cancer.
- **b.** To cover a broad range of topics, including genetics, epigenetics, signal transduction, genomics, proteomics, and metabolomics.
- **c.** To cover the use of various molecular techniques for cancer diagnosis, treatment, and prevention. The course will include lectures, laboratory sessions, and independent study.

d. Intended Learning Outcomes of Course (ILOs):

a- Knowledge and Understanding:

By the end of the course, student should be able to :

1. Identify the key concepts and terminology related to cancer biology and molecular oncology

2. Describe the molecular mechanisms underlying oncogenesis, including genetic and epigenetic alterations

3.Outline the molecular pathways involved in tumor progression and metastasis

Analyze the latest advances in cancer research and emerging trends in the field

4. Underline the potential impact of molecular oncology research on clinical practice and public health

5. Describe a comprehensive understanding of the molecular mechanisms that underlie the development and progression of cancer.

6. Define the role of genetics, epigenetics, signal transduction, genomics, proteomics, and metabolomics in cancer biology.

7. Understand the principles and techniques used in cancer diagnosis, treatment, and prevention.

b- Intellectual Skills:

By the end of the course, student should be able to :

1. Evaluate critically scientific literature related to cancer biology.

2. Design and carry out experiments to investigate the molecular mechanisms of cancer.

6 Page **2** of

3. Analyze and interpret complex data sets related to cancer biology.

c- Professional and Practical Skills:

By the end of the course, student should be able to :

1. Demonstrate proficiency in laboratory techniques used in molecular oncology research

2. Illustrate effective communication skills to convey scientific concepts to both scientific and nonscientific audiences

3. Summarize the ethical issues related to cancer research and its application to clinical practice

d- General Skills:

By the end of the course, student should be able to:

1. Enhance effective time-management and organization skills

2. Develop effective teamwork and collaboration skills

3. Develop critical thinking and problem-solving skills to analyze complex biological problems

4- Course Contents:

Tonia	Hours	Lootunos	Lab.	Practical/
Торіс	nours	Lectures		small groups
Introduction to Cancer Biology:				
Oncogenes and Tumor Suppressor Genes:				
Genetic and Epigenetic Alterations in Cancer:				
DNA Damage and Repair:				
Cell Signaling Pathways in Cancer:				
Tumor Microenvironment:				
Cancer Immunology and Immunotherapy:				
Molecular Diagnostics and Imaging in				
Cancer:				
Cancer Therapeutics:				
Precision Medicine:				
Cancer Stem Cells:				
Cancer Genomics:				
Translational Cancer Research:				
Cancer Epidemiology:				
Cancer Prevention				
Emerging Trends in Cancer Research:				
Total				

:Learning Methods .5

- a. Lectures
- b. Tutorial
- c. Lab.

6. Assessment methods:

	Assessment Type	Date	%
1.	First assessment	16 th week of first term.	30
2.	Lab. Exam	Before the final examination	20
3.	Final examination (Written,)	At the end of term	50
	Т	`otal	100

7. List of References:

Title	Publisher	сору	authors	Available place
The biology of	Garland		Weinberg, R.	
cancer.	Science.		А.	
Hallmarks of			Hanahan, D.,	
cancer: the next			& Weinberg,	
generation. Cell,			R. A.	
Cellular and	Elsevier.	10th ed.	Abbas, A. K.,	
Molecular			Lichtman, A.	
Immunology			H., & Pillai, S.	

8. Facilities Required for Teaching and Learning:

	Facilities Required	Notes
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Course Coordinators

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Head of Department

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The name of university:
Program name
The course name:
Course code:
Academic year / level:

University of Benghazi Molecular diagnostics Biomedical Ethics and Scientific Integrity BMSC-401 2023-2024. Fourth year

5 Page 1 of

1- Basic information:

the course name:	Biomedical Ethics and Scientific Integrity
Course coordinator	
Program (s) on which the course is given:	Molecular diagnostics
Teaching hours	2 hours/week
Language	English
Academic year / level:	2023-2024. Fourth year
Course approval date	

1.1 hours per week

Lecture: (2 hours/week) Training: (-)

Total: (hours 96)

2- Aims of Course:

a- To introduce students to the fundamental principles of biomedical ethics and scientific integrity, and the importance of these principles in the biomedical .sciences

b-To develop students' critical thinking skills, and their ability to analyze .complex ethical issues that arise in biomedical research and practice c-To promote ethical decision-making skills in students, and to provide them with the tools necessary to navigate ethical dilemmas that may arise in their future careers in the biomedical sciences

3- Intended Learning Outcomes of Course (ILOs):

a- Knowledge and Understanding:

After completing this course, students should be able to demonstrate:

- 1. Understand fundamental principles of biomedical ethics and scientific integrity.
- 2. Describe the ethical frameworks and principles that are commonly used in biomedical research and practice.
- 3. Discuss ethical issues that arise in biomedical research and practice, using a range of ethical frameworks and principles

b- Intellectual Skills:

By the end of the course, student should be able to:

- 1. Evaluate complex ethical issues that arise in biomedical research and practice.
- 2. Plan well-reasoned arguments to support ethical decision-making.

c- Professional and Practical Skills:

By the end of the course, student should be able to:

1. Interpret scientific reports and communicate findings to colleagues.

5 Page 2 of

d- General Skills:

By the end of the course, student should be able to:

- 1. Develop skills in teamwork and collaboration.
- 2. Improve problem-solving skills.
- 3. Develop time management skills.
- 4. Develop critical skill required in biomedical research and clinical practice.

4- Course Contents:

Tonic	Hours	Lectures	Lab.	Practical/
Торк	IIOUIS	Lettures		small groups
Introduction to Biomedical Ethics and				
Scientific Integrity				
Autonomy and Informed Consent				
Beneficence and Non-Maleficence				
Justice and Fairness				
Confidentiality and Privacy				
End-of-Life Issues				
Animal Research Ethics				
Ethics of Genetic Testing and Gene Editing				
Clinical Trials				
Organ Donation and Transplantation				
Public Health Ethics				
Reproductive Ethics				
Disability Ethics				
End-of-Life Ethics and Decision-Making				
Technology and Privacy Ethics				
Ethics of Innovation and Entrepreneurship in				
Biomedicine				
Introduction to Scientific Integrity				
Research Design and Data Collection				
Authorship and Publication				
Data Analysis and Interpretation				
Collaboration and Mentoring				
Conflicts of Interest and Professional				
Responsibility				
Scientific Integrity and Research				
Misconduct				
Scientific Integrity and Society				
Research Ethics				
International Perspectives on Scientific				
Integrity				
Total.				

5 Page 3 of
5. Learning Methods:

- a. Lectures
- b. Tutorial
- c. Lab.

6. Assessment methods:

	Assessment Type	Date	%
1.	First assessment	16 th week of first term.	<mark>30</mark>
2.	Lab. Exam	Before the final examination	<mark>20</mark>
3.	Final examination (Written,)	At the end of term	<mark>50</mark>
	100		

7. List of References:

Title	Publisher	сору	authors	Available
				place
Principles of Biomedical Ethics.	New York	8th ed.	Beauchamp,	
	Oxford		T.L. and	
	University Press.		Childress,	
			J.F.	
National Academy of Sciences,	Washington,	3rd ed.		
National Academy of	DC:The National			
Engineering, and Institute of	Academies Press.			
Medicine. On Being a Scientist: A				
Guide to Responsible Conduct in				
Research.				

8. Facilities Required for Teaching and Learning:

	Facilities Required	Notes
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Course Coordinators

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Date:...../...../......

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d. General&	c. Professional &	b. Intellectual skills	Understanding	الدراسد
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Head of Department

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The name of university: Programme name: The course name: Course code: Academic year / level:

University of Benghazi Molecular diagnostics Infection Control and Safety BMSC-402 2023-2024. Fourth year

1. Basic information:

the course name:	Infection Control and Safety
Course coordinator	
Programme (s) on which the course is given:	Molecular diagnostics
Teaching hours	2 hours/week
Language	English
Academic year / level:	2023-2024. Fourth year
Course approval date	

1.1 hours per week

Lecture: (2 hours/week)	Practical: (-)	Training: (-)	Total: (hours 64)
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2. Aims of Course:

- a. To provide students with an understanding of the principles of infection control and safety in healthcare settings.
- b. to equip students with the knowledge and skills necessary to prevent and control infections, protect themselves and patients from hazards, and promote safe and healthy work environments.
- c. to cover a range of topics, including infection control practices, occupational health and safety, emergency preparedness, and risk assessment.

3. Intended Learning Outcomes of Course (ILOs):

a- Knowledge and Understanding:

By the end of the course, student should be able to :

a1. Reproduce a comprehensive understanding of the principles of infection control and safety.

- a2. Identify the different types of infections and their modes of transmission.
- a3. State the role of healthcare professionals in preventing and controlling infections.
- a4. Describe the principles of occupational health and safety in healthcare settings.
- a5. Outline the principles of emergency preparedness in healthcare settings.

b- Intellectual Skills:

By the end of the course, student should be able to :

- b1. Evaluate the effectiveness of infection control practices in healthcare settings.
- b2. Analyze and assess the risks associated with healthcare practices.
- b3. Develop strategies to prevent and control infections in healthcare settings.
- b4. Develop emergency preparedness plans for healthcare settings.
- b5. Interpret and analyze infection control policies and procedures.

c- Professional and Practical Skills:

By the end of the course, student should be able to :

1. Apply infection control practices in healthcare settings.

- 2. Use appropriate personal protective equipment in healthcare settings.
- 3. Assess risk assessments in healthcare settings.
- 4. Illustrate and implement infection control policies and procedures.
- 5. Operate effectively with colleagues and patients about infection control practices.

d- General Skills:

- By the end of the course, student should be able to:
- d1. Develop critical thinking skills.
- d2. Work effectively in teams.
- d3. Develop problem-solving skills.
- d4. Improve time management skills.
- d5. Develop effective communication skills.

4- Course Contents:

Торіс	Hours	Lectures	Lab.	Practical/
				small groups
Introduction to Infection Control and Safety				
Infection Control Practices				
Personal Protective Equipment (PPE)				
Hand Hygiene				
Environmental Cleaning and Disinfection				
Sterilization and Disinfection of Medical				
Devices				
Infection Prevention and Control in Special				
Situations				
Occupational Health and Safety				
Emergency Preparedness				
Risk Assessment				
Healthcare-Associated Infections (HAIs)				
Outbreak Management				
Communicable Diseases				

Tonic	Hours	Lectures	Lab.	Practical/
Торк	Hours	Lettures		small groups
Immunization and Vaccination				
Infectious Waste Management				
Emerging Infectious Diseases				
Infection Control Policies and Procedures				
Infection Control Auditing and Surveillance				
Management of Infectious Diseases				
Infection Control in the Community				
Infection Control in Outpatient Settings				
Infection Control in Long-Term Care Facilities				
Infection Control in the Laboratory				
Infection Control Program Evaluation				
Infection Control in a Pandemic				
Total				

5. Learning Methods:

- a. Lectures
- b. Tutorial

6. Assessment methods:

	Assessment Type	Date	%
1.	First assessment	16 th week of first term.	40
2.	Final examination (Written,)	At the end of term	60
	100		

7. List of References:

Title	Publisher	сору	authors	Available place

8. Facilities Required for Teaching and Learning:

	Facilities Required	Notes
1.		
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Course Coordinators

Head of Department

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Date:...../...../....../

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	Skills												a.				الاسبوع			
t	d. General& transferable skills					e. Pro prac	fessio tical	onal d skills	& 8	b. Intellectual skills						Knowledge and Understandi ng				الدراسد <i>ي</i>
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The name of university:
Programme name:
The course name:
Course code:
Academic year / level:

University of Benghazi Molecular diagnostics Molecular Diagnostic of Infectious Diseases MOLD-403 2023-2024. Fourth year

1. Basic information:

the course name:	Molecular Diagnostic of Infectious Diseases
Course coordinator	
Programme (s) on which the course is given:	Molecular diagnostics
Teaching hours	5 hours/week
Language	English
Academic year / level:	2023-2024. Fourth year
Course approval date	

1.1 hours per week

Lecture: (3 hours/week)

Practical: (2 hours/week) Training: (-) Total: (hours 160)

2. Aims of Course:

- a. To equip students with the necessary knowledge and practical skills required to work in a clinical laboratory
- b. to make informed decisions regarding the selection and interpretation of appropriate diagnostic tests for the diagnosis and management of infectious diseases.

3. Intended Learning Outcomes of Course (ILOs):

a- Knowledge and Understanding:

By the end of the course, student should be able to :

a1. Define the principles and concepts of molecular diagnostics of infectious diseases.

a2. Outline the different types of molecular techniques used in the diagnosis of infectious diseases.

a3. Identify the interpretation of molecular diagnostic results.

a4. Understand the principles and techniques used in molecular diagnostics of infectious diseases.

a5. Describe the various molecular diagnostic methods used to identify and diagnose infectious diseases.

a6. Define the clinical applications of molecular diagnostics in infectious diseases.

a7. Analyze and interpret molecular diagnostic results in clinical setting.

b- Intellectual Skills:

By the end of the course, student should be able to :

b1. Adapt critical thinking and problem-solving skills to molecular diagnostic techniques and their application in the diagnosis of infectious diseases.

b2. Analyze and interpret molecular diagnostic results in a clinical setting.

b3. Evaluate the performance of diagnostic tests and make informed decisions regarding the selection and interpretation of appropriate tests.

c- Professional and Practical Skills:

By the end of the course, student should be able to :

c1. Demonstrate practical skills in molecular diagnostic techniques used in the diagnosis of infectious diseases.

- c2. Prepare and apply experimental protocols to optimize molecular diagnostic methods.
- c3. Operate collaboratively in a laboratory environment.

d- General Skills:

By the end of the course, student should be able to:

- d1. Communicate effectively through written and oral presentations.
- d2. Analyze and interpret scientific data.

4- Course Contents:

Торіс	Hours	Lectures	Lab.	Practical/
· r ·				small groups
Introduction to Molecular Diagnostics				
Nucleic Acid Extraction Techniques				
Polymerase Chain Reaction (PCR)				
Real-Time PCR				
Reverse Transcriptase PCR (RT-PCR)				
Loop-Mediated Isothermal Amplification (LAMP)				
Next-Generation Sequencing (NGS)				
Microarray Technology				
Multiplex PCR				
Digital PCR				
Hybridization-Based Techniques				
MALDI-TOF Mass Spectrometry				
Infectious Disease Diagnostics in Developing Countries				
Clinical Applications of Molecular Diagnostics				

Торіс	Hours	Lectures	Lab.	Practical/ small groups			
Emerging Infectious Diseases							
Pathogen Genomics							
Total							

5. Learning Methods:

- a. Lectures
- b. Tutorial
- c. Lab.

6. Assessment methods:

	Assessment Type	Date	%					
1.	First assessment	16 th week of first term.	30					
2.	Lab. Exam	Before the final examination	20					
3.	Final examination (Written,)	At the end of term	50					
	Total							

7. List of References:

Title	Publisher	сору	authors	Available place
Real-time PCR in	Nucleic acids		Mackay IM,	
virology	research		Arden KE,	
			Nitsche	
Tolerance of loop-	Journal of		Kaneko H,	
mediated isothermal	bioscience and		Kawana T,	
amplification to a	bioengineering.		Fukushima E,	
culture medium and			Suzutani T.	
biological				
substances				

8. Facilities Required for Teaching and Learning:

	Facilities Required	Notes
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Course Coordinators

Head of Department

7 Page 4 of

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Date:...../...../....../

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The name of university: Programme name: The course name: Course code: Academic year / level: University of Benghazi Molecular diagnostics Molecular Therapeutics MOLD-405 2023-2024. Fourth year

7 Page 1 of

1. Basic information:

the course name:	Diagnostic Genetics
Course coordinator	
Programme (s) on which the course is given:	Molecular diagnostics
Teaching hours	5 hours/week
Language	English
Academic year / level:	2023-2024. Fourth year
Course approval date	

1.1 hours per week

Lecture: (3 hours/week)

Practical: (2 hours/week) Training: (-) Total: (hours 160)

2. Aims of Course:

a. To provide students with a comprehensive understanding of the principles and practices of molecular therapeutics, including the use of genetic, molecular, and cellular approaches to drug development, the optimization of drug efficacy and safety, and the regulatory and ethical considerations of drug development.

3. Intended Learning Outcomes of Course (ILOs):

a- Knowledge and Understanding:

By the end of the course, student should be able to :

- a1. Define the principles of molecular biology and their relevance to medicine
- a2. Recognize the techniques involved in the development of molecular therapeutics
- a3. Describe the different types of molecular targets for drug development
- a4. Identify the role of genomics and proteomics in drug discovery

b- Intellectual Skills:

By the end of the course, student should be able to :

- b1. Evaluate critically the scientific literature related to molecular therapeutics
- b2. Design and develop molecular therapeutics based on scientific principles

c- Professional and Practical Skills:

By the end of the course, student should be able to :

c1. Apply molecular biology techniques in drug discovery and development

c2. Communicate scientific information effectively to both scientific and non-scientific audiences

c3. <u>Illustrate</u> the regulatory and ethical issues associated with the development and use of molecular therapeutics

d- General Skills:

By the end of the course, student should be able to:

- d1. **Develop** critical thinking and problem-solving
- d2. **<u>Develop</u>** effective communication

7 Page 2 of

- d3. <u>Develop</u> Teamwork and collaboration
- d4. **Develop** Time management and organization.

4- Course Contents:

Tonio	Hours	Lootunos	Lab.	Practical/
Topic	nours	Lectures		small groups
Introduction				
Simple protein replacement therapy				
Recombinant protein production				
Recombinant vaccines				
Therapeutic antibodies and immunotherapy				
Transgenic animals				
Transplantation: a form of gene therapy				
Reproductive cloning				
Stem cell therapy				
Gene augmentation therapy				
Gene therapy trials for inherited diseases				
Gene silencing technologies				
Gene therapy for cancer				
Single-nucleotide polymorphisms (SNPs)				
and therapy				
Legislation, clinical trials and ethical issues				
Total				

5. Learning Methods:

- a. Lectures
- b. Tutorial
- c. Lab.

6. Assessment methods:

	Assessment Type	Date	%
1.	First assessment	16 th week of first term.	30
2.	Lab. Exam	Before the final examination	20
3.	Final examination (Written,)	At the end of term	50
	Т	otal	100

7. List of References:

Title	Publisher	сору	authors	Available place
Principles of	Dan Longo	Second	J. Larry	
Molecular Medicine			Jameson,	
			Joseph	
			Loscalzo,	

7 Page 3 of

		Dennis Kasper,	
		Stephen	
		Hauser	
Molecular Medicine		Jens Kurreck,	
		Tanja Weil,	
		and Stefan	
		Endres.	
Molecular	Springer	Greenwell, P.,	
Therapeutics		& McCulley,	
		M.	

8. Facilities Required for Teaching and Learning:

	Facilities Required	Notes
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Course Coordinators

Head of Department

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Date:...../...../......

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The name of university: Programme name: The course name: Course code: Academic year / level: University of Benghazi Molecular diagnostics Seminar in Molecular Diagnostics MOLD-408 2023-2024. Fourth year

1. Basic information:

the course name:	Seminar in Molecular Diagnostics
Course coordinator	
Programme (s) on which the course is given:	Molecular diagnostics
Teaching hours	1 hours/week
Language	English
Academic year / level:	2023-2024. Fourth year
Course approval date	

1.1 hours per week

Lecture: (1 hours/week)	Practical: (-)	Training: (-)	Total: (hours 32)
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2. Aims of Course:

- a. To develop students' critical thinking and presentation skills in the context of molecular diagnostics.
- b. To explore and present on a variety of topics related to the application of molecular diagnostic techniques in biomedical research and clinical practice.
- c. To promote critical thinking, effective communication, and research skills.
- d. To learn about new topics, share knowledge and ideas, and improve public speaking and presentation skills.

3. Intended Learning Outcomes of Course (ILOs):

a- Knowledge and Understanding:

By the end of the course, student should be able to :

a1. <u>Describe</u> the principles of molecular biology and how they relate to disease diagnosis.

a2. <u>Outline</u> the basics of DNA sequencing, PCR, gene expression analysis, and next-generation sequencing.

a3. **Define** the applications of molecular diagnostics in clinical and research settings.

b- Intellectual Skills:

By the end of the course, student should be able to :

- b1. Analyze and evaluate critically molecular diagnostic data.
- b2. Synthesize and interpret molecular diagnostic data to formulate a diagnosis.
- b3. Develop critical thinking and problem-solving skills.

c- Professional and Practical Skills:

By the end of the course, student should be able to :

- c1. Demonstrate proficiency in public speaking and presentation skills.
- c2. Interpret laboratory results and communicate findings effectively.
- c3. Develop skills in conducting literature reviews and presenting scientific information.

d- General Skills:

By the end of the course, student should be able to:

5 Page 2 of

- d1. **<u>Develop</u>** written and oral communication skills.
- d2. Develop teamwork and collaboration skills.
- d3. Develop time management and organizational skills.

4- Course Contents:

Tonia	Hours	Locturos	Lab.	Practical/
Topic	mours	Lectures		small groups
Introduction to molecular diagnostics.				
Principles of molecular biology.				
DNA sequencing technologies and .applications				
Polymerase chain reaction (PCR) techniques and applications.				
Gene expression analysis.				
Next-generation sequencing technologies and applications.				
Microarray technology and applications.				
Molecular diagnostics in cancer.				
Molecular diagnostics in infectious diseases.				
Molecular diagnostics in genetic disorders.				
Quality control and assurance in molecular diagnostics.				
Ethical considerations in molecular diagnostics.				
Applications of molecular diagnostics in personalized medicine.				
Trends and future directions in molecular diagnostics.				
Oral and written presentation skills in molecular diagnostics.				
Total				

5. Learning Methods:

- a. Lectures
- b. Tutorial

6. Assessment methods:

	Assessment Type	Date	%
1.	First assessment	16 th week of first term.	40
2.	Final examination (Written,)	At the end of term	60
	100		

7. List of References:

Title	Publisher	сору	authors	Available place			

8. Facilities Required for Teaching and Learning:

	Facilities Required	Notes
1.		
2.		
3.		

Course Coordinators

Head of Department

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Date:...../...../....../

Skills										a Knowledge and				الاسبوع						
d. General& transferable skills				c. Professional & practical skills				b. Intellectual skills					Understanding				الدراسي			
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